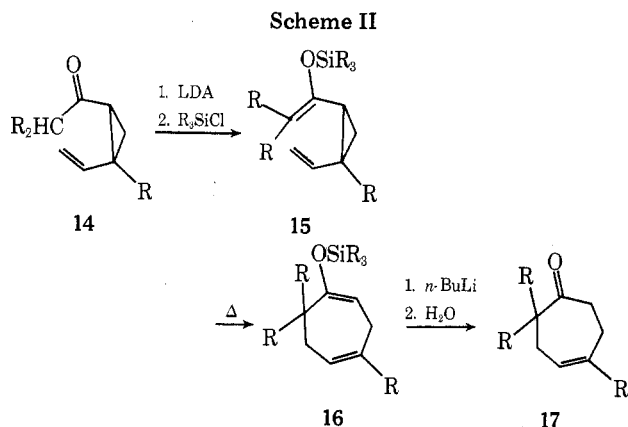


A typical procedure for the above sequence is described below for the conversion of alkoxyenone **1** to ketone **4**. To a -78°C ether (80 ml) solution of 1-bromo-2-vinylcyclopropane (40 mmol, 7:3 mixture of *cis* and *trans*) was added *t*-BuLi (48 mmol, 1.5 M in pentane) over 5 min. The resulting solution was stirred for 1.5 h at -78°C and subsequently warmed to 0°C . An ether (10 ml) solution of alkoxyenone **1** (20 mmol) was then added over a period of 2 min. The reaction mixture was stirred for 15 min at 0°C and 15 min at ambient temperature and then carefully poured into a separatory funnel containing 2 N HCl (100 ml). Intermittent agitation of the above mixture (15 min) followed by standard workup provided in 91% yield (90% purity) the divinylcyclopropanes **3a** and **3b** (7:3 mixture of *cis* and *trans*), which were purified by silica gel chromatography (ether-hexane, 3:7).⁷ Compounds **3a** and **3b** (2 M benzene solution) upon thermolysis (170 – 180°C , 2 h) in a sealed Pyrex tube provided after purification ketone **4** (bp 54 – 59°C , 0.15 mm) in 87% yield. Alternatively ketone **4** can be prepared by heating (250°C , 5 min) the crude mixture of compounds **3a** and **3b** in a distillation apparatus followed directly by distillation of ketone **4**. In either case the method offers an exceptionally straightforward route to the annelated product with an overall yield of 72%.

The above strategy could be readily extended to other cycloheptane systems by varying the starting substrate (latent double bond equivalent). For example, aldehydes could be readily converted to acylvinylcyclopropanes **14** which could be used in the preparation of cyclohept-4-enones as outlined in Scheme II.



In order to test the efficacy of this route to cycloheptenones and examine variations in the lithio reagents, we have investigated this strategy in an approach to karahanaenone (17, R = Me).¹⁰ Thus, a mixture of *cis*- and *trans*-1-lithio-2-methyl-2-vinylcyclopropane,¹¹ obtained from the metallation of the corresponding bromides, upon reaction with isobutyraldehyde and oxidation (pyridinium chlorochromate)¹² of the resulting alcohols provided a mixture of ketones **14** (R = Me, *cis* and *trans*). Treatment of this mixture with lithium diisopropylamide (LDA) in tetrahydrofuran (THF) followed by quenching with trimethylsilyl chloride afforded the silyloxy-divinylcyclopropanes **15** (R = Me, *cis* and *trans*). Thermolysis (165 – 175°C , 1.5 M benzene solution) of this mixture followed by desilylation (*n*-BuLi, THF, 25°C , 5 h) of the resulting diene **16** (R = Me) provided karahanaenone (**17**, R = Me) in an overall yield of 54% based on isobutyraldehyde.

Studies on the preparation of more highly functionalized reagents and the application of these reagents in synthesis are in progress.

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References and Notes

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- (3) The pure *cis*- and *trans*-1-bromo-2-vinylcyclopropanes were obtained in preparative quantities by spinning-band distillation of the mixture of bromides prepared from butadiene. For the preparation of the bromides, see L. Skattebøl, *J. Org. Chem.*, **29**, 2951 (1964), and D. Seyferth, H. Yamazaki, and D. L. Alleston, *ibid.*, **28**, 703 (1963). For stereochemical assignments, see J. Landgrebe and L. Becker, *ibid.*, **33**, 1173 (1968).
- (4) Compound **3a**: ir (neat) 1695 and 1635 cm^{-1} ; NMR (CCl_4) δ 0.80–2.4 (complex m with br s at 1.71, 1.1 H) and 4.80–5.68 (m, 3 H).
- (5) All new compounds reported were homogeneous by TLC or VPC and gave satisfactory ir and NMR spectra and exact mass or combustion analyses.
- (6) Compound **3b**: ir (neat) 1695 and 1635 cm^{-1} ; NMR (CCl_4) δ 0.80–2.60 (complex m with br s at 1.71, 1.1 H) and 4.80–5.50 (m, 3 H).
- (7) Alternatively the divinylcyclopropanes may be isolated by distillation in which case the distillate is contaminated with varying amounts of the annelated product depending on the temperature used for the distillation. Pure samples of **3b** obtained by silica gel chromatography, upon standing at ambient temperature, slowly rearranged into compound **4**. In the other cases reported in Table I isolation of the intermediate *cis*-divinylcyclopropanes can only be accomplished by low temperature chromatography owing to significant rearrangement at room temperature.
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- (9) Compound **4**: bp 54 – 59°C (0.15 mm); ir (neat) 1740 cm^{-1} ; NMR (CCl_4) δ 1.09 (s, 3 H), 2.05–3.10 (complex m, 8 H), and 5.30–5.77 (m, 3 H).
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- (13) Dreyfus Foundation Fellow, 1975–1976.

Paul A. Wender,* Michael P. Filosa¹³

Department of Chemistry, Harvard University
Cambridge, Massachusetts 02138

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Alkylation and Michael Additions of Glycine Ethyl Ester. Use in α -Amino Acid Synthesis and as Acyl Carbanion Equivalent

Summary: The benzylidene derivative of glycine ethyl ester can be used in mono- or sequential dialkylations thus leading to very simple syntheses of α -amino esters and acids; michael addition can also be effected readily, especially in protic solvents; the α -amino ester functionality can be transformed into a carbonyl (lithium aluminum hydride; periodate) and glycine ethyl ester is thus an acyl carbanion equivalent.

Sir: We would like to report that the readily available benzylidene derivatives of glycine esters can be alkylated in high yield under a variety of conditions. This obviously provides a particularly simple route to α -amino acids.¹

It is especially noteworthy that the relatively high acidity of **1** permits formation of the anion and its alkylation not only with strong bases like lithium diisopropylamide, but with weaker bases such as potassium *tert*-butoxide. Also noteworthy is the fact that these alkylations can be performed not only with the *tert*-butyl ester, but are very satisfactory with the simple ethyl ester, in spite of the "extreme instability" claimed for this substance.² Because an α -amino ester is a masked carbonyl group, the anion of a benzylidene glycine ester is also an acyl carbanion equivalent.³ The latent carbonyl function may be unmasked, *inter alia*, via the sequence lithium

- (2) Prepared by treating 10.1 g of glycine ethyl ester hydrochloride in 150 ml of methylene chloride with 1 equiv of benzaldehyde in the presence of 20 ml of triethylamine and 6 g of anhydrous magnesium sulfate at room temperature, filtration, solvent removal (room temperature), water-ether partition washing (brine), drying, and removal of solvent. The substance thus obtained in 95% yield could be kept in the freezer for several months. See also O. Gerngross and A. Olcay, *Ber.*, **96**, 2550 (1963).
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- (6) All new compounds gave integrated NMR spectra in complete agreement with their structures which were also confirmed by their further transformations. Kugelrohr distillations are indicated as bp (k).
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- (12) On leave from the Université Pierre et Marie Curie, Paris.

Gilbert Stork,* Ambrose Y. W. Leong
Anne Marie Touzin¹²

Department of Chemistry, Columbia University
New York, New York 10027

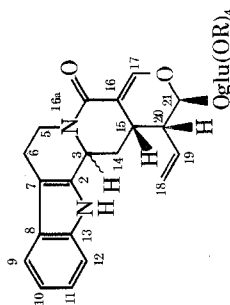
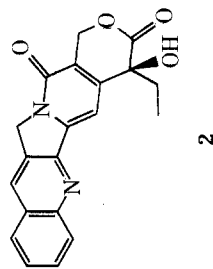
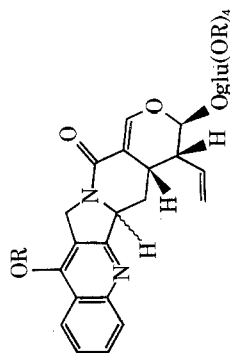
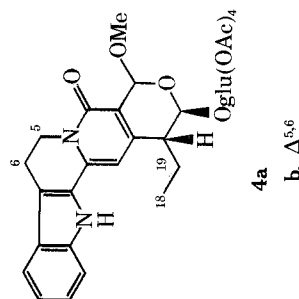
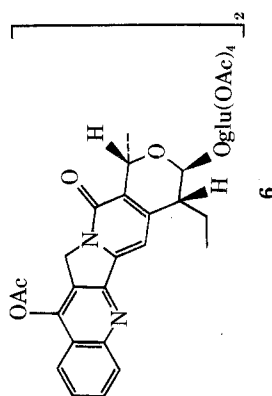
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A Biomimetic Synthesis of the Camptothecin Chromophore

Summary: Novel heterocyclic alkaloids (4 and 6), potential synthetic precursors of 20(S)-camptothecin (2), are synthesized by 2,3-dichloro-5,6-dicyanobenzoquinone oxidation of tetraacetyl-18,19-dihydrovincoside (18,19-H₂-1a) and -isovincoside (18,19-H₂-1c) lactams and their corresponding pentaacetyl-18,19-dihydroquinolols (18,19-H₂-3).

Sir: We have been studying the chemistry¹ of the penultimate biosynthetic precursor of camptothecin (2), isovincoside lactam (1c), as a model system for the putative biochemical transformations that occur between 1c and 2 in vivo.² Since D ring oxidation of 1c to a pyridone may be one requisite of the biosynthetic pathway to 2, we have examined the oxidation of 18,19-H₂-1a and -1c using 2,3-dichloro-5,6-dicyanobenzoquinone (DDQ). Alternatively, D ring oxidation of isovincoside quinolol (3c) may be a key oxidative step preceding 2 in vivo, since presently we do not know the exact biochemical sequence of events between 1c and 2.³ With both 1 and 3 oxidation with DDQ has been accomplished efficiently, which should enable a convenient synthesis of 2 and novel indole analogues of it, and which may be relevant to in vivo biosynthetic events.⁴

Oxidation of either 18,19-H₂-1b or -1d (OAc)₄ with DDQ (1 equiv or excess) in methanol (reflux, 5 min, N₂) or in a toluene-methanol mixture (25 °C, 5-10 min, N₂) gave a chromatographically resolvable mixture of 4a [pale yellow solid: mp 145-150 °C dec; 41%; ν_{KBr} 3356 (NH), 1761 (OAc), 1667 (pyridone), and 1230 (C-O) cm⁻¹; $\text{uv } \lambda_{\text{max}}^{\text{MeOH}}$ 386, 367, 296 (sh), 286 (sh), 273, 260, 252, and 213 nm; MS *m/e* 666.2437 (M⁺ - CH₂O, calcd for C₃₄H₃₈N₂O₁₂ 666.2424), 331.1026 [Glu(OAc)₄⁺, calcd for C₁₄H₁₉O₉ 331.1024]; ¹H NMR (90 MHz) δ^{CDCl_3} 0.93 [t, 3 H, *J* = 7 Hz, C(18)], 1.89 [m, 2 H, C(19)], 2.00-2.07 (4 s, 12 H, 4 OAc), 2.58 [m, 1 H, C(20)], 2.95 [t, 2 H, *J* = 7 Hz, C(6)], 3.56 (s, 3 H, OCH₃), 4.35 [t, 2 H, *J* = 7 Hz, C(5)], 5.41 (d, 1 H, *J* = 3 Hz, C(21)], 5.70 [s, 1 H, C(17)], 6.32 (s, 1 H, C(14)], 7.08-7.54 (4 aromatic H), and 9.51 (br s, NH), glucosyl protons omitted] and 4b [yellow needles (MeOH); mp 154-156.5 °C; 26.5%; ν_{KBr} 3333 (NH), 1754 (OAc), 1658 (pyridone), and 1230 (C-O) cm⁻¹; $\text{uv } \lambda_{\text{max}}^{\text{EtOH}}$ 418, 395 (sh), 324, 277, 257, 248 (sh), and 218 nm; MS *m/e* 664.1897 (M⁺ - CH₂O, calcd for C₃₄H₃₆N₂O₁₂ 664.2258), 316.1153 [M⁺ + 1 - CH₃O - (HO-Glu(OAc)₄, calcd for C₂₀H₁₆N₂O₂ 316.1027], and 290.1419 (calcd for C₁₉H₁₈N₂O); ¹H NMR (90 MHz) δ^{CDCl_3} 1.02 [t, 3 H, *J* = 7 Hz, C(18)], 1.90 [m, 2 H, C(19)], 1.96-2.07 (4 s, 12 H, 4 OAc), 2.90 [m, 1 H, C(20)],



- 1a, C-3 (R), R = H
- 1b, C-3 (R), R = Ac
- 1c, C-3 (S), R = H
- 1d, C-3 (S), R = Ac
- 3a, C-3 (R), R = H
- 3b, C-3 (R), R = Ac
- 3c, C-3 (S), R = H
- 3d, C-3 (S), R = Ac
- 4a, $\Delta^{5,6}$
- 4b, $\Delta^{5,6}$

Table I. ¹³C NMR Assignments^a

Compd	Carbon resonances at 22.6 MHz ^b																
	18	19	6	5	20	OMe	21	14	17	12	16	10	9	11	8	16a	
4a	10.4	19.5	22.7	40.6	42.6	56.2	96.0	96.5	100.1	114.6	119.6	120.2	124.6	125.7	127.2	138.0	138.7
4b	11.1	21.1	111.9	108.8	43.8	56.3	96.1	96.1	98.1	110.6	118.6	120.5 ^c	126.7	120.7 ^c	129.9	139.8	132.9
6	9.4	20.8	123.9	48.8	42.0	98.5	99.4	89.7	89.7	130.8	120.1	122.3	127.9 ^d	129.6 ^d	121.3	150.8	150.8
																145.1	149.4
																146.7	148.3
																160.4	160.4

^a Determined in CDCl₃ relative to TMS as internal standard by consideration of the ¹³C NMR assignments of 1b and 1d and 2, ⁷SFOR proton decoupling, and low power proton decoupling. ^b Glucosyl tetraacetate resonances omitted. ^c These two assignments may be reversed. ^d These two assignments could be interchanged.